

AD-A193 875

NEGATIVE CONTROL OF BIODEGRADATION IN PSEUDOMONAS(U)  
YALE UNIV NEW HAVEN CONN L N ORNSTON MAR 88  
ARO-21256.3-LS DAAG29-84-K-0151

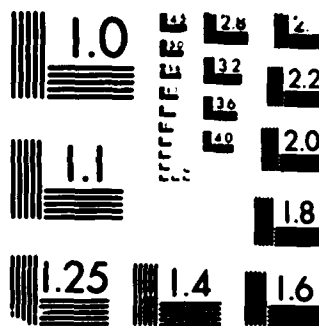
1/1

UNCLASSIFIED

F/G 6/2

NL





MICROCOPY RESOLUTION TEST CHART  
101-A

AD-A193 875

UNCLASSIFIED  
SECURITY CLASSIFICATION OF THIS PAGE

MASTER COPY

FOR REPRODUCTION PURPOSES

2

DTIC FILE COPY			REPORT DOCUMENTATION PAGE			
1a. REPORT SECURITY CLASSIFICATION Unclassified			1b. RESTRICTIVE MARKINGS			
2a. SECURITY CLASSIFICATION AUTHORITY			3. DISTRIBUTION/AVAILABILITY OF REPORT Approved for public release; distribution unlimited.			
2b. DECLASSIFICATION/DOWNGRADING SCHEDULE APR 13 1988			5. MONITORING ORGANIZATION REPORT NUMBER(S) ARO 21256.3-LS			
4. PERFORMING ORGANIZATION REPORT NUMBER(S)			7a. NAME OF MONITORING ORGANIZATION U. S. Army Research Office			
6a. NAME OF PERFORMING ORGANIZATION Yale University			6b. OFFICE SYMBOL (If applicable)		7b. ADDRESS (City, State, and Zip Code) P. O. Box 12211 Research Triangle Park, NC 27709-2211	
8a. NAME OF FUNDING/SPONSORING ORGANIZATION U. S. Army Research Office			8b. OFFICE SYMBOL (If applicable)		9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER DAAG29-84-K-0151	
8c. ADDRESS (City, State, and Zip Code) P. O. Box 12211 Research Triangle Park, NC 27709-2211			10. SOURCE OF FUNDING NUMBERS PROGRAM ELEMENT NO. PROJECT NO. TASK NO. WORK UNIT ACCESSION NO.			
11. TITLE (Include Security Classification) Negative Control of Biodegradation in Pseudomonas						
12. PERSONAL AUTHOR(S) L. Nicholas Ornston						
13a. TYPE OF REPORT Final		13b. TIME COVERED FROM 1/1/85 TO 2/31/87		14. DATE OF REPORT (Year, Month, Day) March 1988		15. PAGE COUNT 1
16. SUPPLEMENTARY NOTATION The view, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy, or decision, unless so designated by other documentation.						
17. COSATI CODES FIELD GROUP SUB-GROUP			18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number) Bacteria, Aromatic Catabolism, Biodegradation, Pseudomonas, Enzymes			
19. ABSTRACT The objective of the research program was to characterize control mechanisms that exercised negative regulation on the expression of genes for aromatic catabolism in bacteria. The three sets of genes selected for investigation were <i>ben</i> , encoding enzymes that convert benzoate to catechol; <i>cat</i> , encoding enzymes that convert catechol to citric acid cycle intermediates; and <i>pca</i> , encoding enzymes that convert protocatechuate to citric acid cycle intermediates. Our initial approach was to clone the structural genes from <i>Acinetobacter calcoaceticus</i> and <i>Pseudomonas putida</i> , bacteria in which aromatic catabolism has been well characterized, because we knew that regulatory genes frequently flanked the structural genes.  Our efforts were largely successful, and we identified two cloned regulatory genes from <i>P. putida</i> . One of these, <i>pcaR</i> , exercises positive control over three unlinked gene clusters. The other, <i>catR</i> , exercises negative control over the tightly linked <i>catBC</i> genes. The latter gene is analogous in many respects to another regulatory gene, also designated <i>catR</i> , that we have cloned from <i>A. calcoaceticus</i> .						
20. DISTRIBUTION STATEMENT/AVAILABILITY OF ABSTRACT <input type="checkbox"/> UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT. <input type="checkbox"/> DTIC USERS				21. ABSTRACT SECURITY CLASSIFICATION Unclassified		
22a. NAME OF RESPONSIBLE INDIVIDUAL				22b. TELEPHONE (Include Area Code)		22c. OFFICE SYMBOL

UNCLASSIFIED

SECURITY CLASSIFICATION OF THIS PAGE

UNCLASSIFIED

SECURITY CLASSIFICATION OF THIS PAGE

## FINAL TECHNICAL REPORT FOR DDAG29-84-K-0151

"Negative Control of Biodegradation in *Pseudomonas*"

The objective of the research program was to characterize control mechanisms that exercised negative regulation on the expression of genes for aromatic catabolism in bacteria. The three sets of genes selected for investigation were *ben*, encoding enzymes that convert benzoate to catechol; *cat*, encoding enzymes that convert catechol to citric acid cycle intermediates; and *pca*, encoding enzymes that convert protocatechuate to citric acid cycle intermediates. Our initial approach was to clone the structural genes from *Acinetobacter calcoaceticus* and *Pseudomonas putida*, bacteria in which aromatic catabolism has been well characterized, because we knew that regulatory genes frequently flanked the structural genes.

Our efforts were largely successful, and we identified two cloned regulatory genes from *P. putida*. One of these, *pcaR*, exercises positive control over three unlinked gene clusters. The other, *catR*, exercises negative control over the tightly linked *catBC* genes. The latter gene is analogous in many respects to another regulatory gene, also designated *catR*, that we have cloned from *A. calcoaceticus*.

Cloned genes from *A. calcoaceticus* have provided the most accessible example of negative control. When expressed under a *lac* promoter in *E. coli*, the *benABC* genes enable the cells to rapidly convert benzoate to benzoate diol which accumulates in the culture broth. When expressed in an *A. calcoaceticus* mutant lacking benzoate diol dehydrogenase, the *benABC* genes are almost inactive. We do not know whether the lack of activity is due to transcriptional control or due to inactivation of gene products, and we intend investigate these alternatives.

Analysis of the *pca* gene order revealed remarkable rearrangements that took place as the *A. calcoaceticus* structural genes diverged from their homologs in *P. putida*. For example, genes from the former species are tightly clustered in the order *pcaBDC* whereas their homologs, also tightly clustered, appear in the order *pcaBCD* in *P. putida*. The *pcaE* gene is clustered with and transcribed with the other *pca* genes in *A. calcoaceticus*. In *P. putida*, the *pcaE* gene is separated by more than 15 kilobase pairs from the other *pca* genes, although it remains under control of the *pcaR* gene.

Additional functions remain to be explored in *P. putida*. Among these are benzoate chemotaxis, also under control of the *pcaR* gene, and ketoadipate transport. The latter function appears to be associated with scavenging of an aromatic catabolite during starvation-survival. High-level futile activity of the transport system is lethal to starved cells, and the study of mutants resistant to this effect may give some insight into physiology of starvation.

<input checked="" type="checkbox"/>
<input type="checkbox"/>
<input type="checkbox"/>
Index
or
1
<div style="border: 1px solid black; border-radius: 50%; padding: 5px; display: inline-block;"> A-1 </div>
<div style="border: 1px solid black; border-radius: 50%; padding: 5px; display: inline-block;"> COPY INSPECTED 4 </div>

END

DATE

FILMED

8-88

DTIC